Laboratory diagnostics for Lyme disease

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Abstract

Background: Lyme disease is on the rise in Canada. It is a notifiable disease, and when infection is disseminated, serological testing provides supplemental evidence to confirm a case.

Objective: To describe the current diagnostic tests for Lyme disease, review the recommended approach to laboratory testing for Lyme disease and identify future research priorities for Lyme disease laboratory diagnostics in Canada.

Methods: A review of the literature was carried out. We then summarized parameters to consider before Lyme disease testing is conducted, described the current best practice to use a two-tiered diagnostic algorithm for the laboratory confirmation of disseminated Lyme disease, and analyzed the advantages and disadvantages of the supplemental tests for Lyme disease.

Results: Diagnostic testing is indicated in people who have symptoms of disseminated disease and a history of exposure to vector ticks. To maximize sensitivity and specificity, a two-tiered serological approach is recommended, consisting of an enzyme immunoassay (EIA) screening test followed by confirmation with Western blot (WB) testing. A number of other diagnostic tests are available; however, these are largely for research purposes.

Conclusion: Two-tiered serology is currently the best approach available to assist doctors when they are making a diagnosis of disseminated Lyme disease. The Public Health Agency of Canada (the Agency) will seek to improve on this approach through standardization of the Lyme disease diagnostics used across laboratories in Canada, evaluation of test performance characteristics of current and new diagnostic platforms and development of a process to secure robust serum panels to assist in the development and evaluation of new diagnostic tests for Lyme disease.

Introduction

Lyme disease (LD) is a tick-borne infection caused primarily by three species of spirochetes in the *Borrelia burgdorferi* sensu lato genogroup: *B. burgdorferi* sensu stricto (in North America and Western Europe), *B. afzelii* (in Western Europe, central Europe and Russia) and *B. garinii* (primarily in Europe, Russia and northern Asia) (1). The symptoms of Lyme disease occur in stages and involve a variety of tissues and organs, including the skin, joints, heart and nervous system (2). There has been a steady increase in the incidence of Lyme disease in parts of central and eastern Canada (3-5) due to the recent range expansion of the primary tick vector, *Ixodes scapularis* (6).

Lyme disease has been a nationally notifiable disease since 2009 (7). The objective of this article is to describe the current diagnostic tests for Lyme disease, including a review of the recommended approaches to laboratory testing, and identify future research priorities for Lyme disease diagnostics in Canada.

Methods

An extensive review of peer-reviewed literature was carried out. We then summarized the key parameters to consider before Lyme disease testing is conducted, described the current best practice of a two-tiered serological algorithm for the laboratory confirmation of disseminated Lyme disease and explored the advantages

and disadvantages of the supplemental tests. We also outlined future research plans to be undertaken by the Agency's National Microbiology Laboratory.

Results

Considerations prior to testing

Early localized Lyme disease does not require diagnostic testing before antibiotic therapy is started. A presumptive diagnosis can be made on the basis of the clinical presentation and a credible history of exposure to infected blacklegged ticks (8). Typically, diagnostic testing is appropriate for people with a history of tick exposure and symptoms of disseminated Lyme disease infection, since test sensitivity improves as the bacteria affect tissue systems other than the skin (8,9). Testing, however, should be limited to those with objective signs of infection (10,11).

The following information is required prior to testing:

- Detailed travel history and date of onset of symptoms This information should be included on the laboratory requisition, as it helps the diagnostic laboratory apply the most appropriate test platform. For example, there are different tests to identify Lyme disease acquired in Europe/Asia versus North America (12), and different tests are used for early infections versus infections that may have been present for some time (13).
- A history of antibiotic treatment This can dampen the immune response to infection and may complicate the interpretation of serological tests (14).
- Other infections or pre-existing conditions Infection with other related pathogens (e.g. syphilis) and autoimmune disorders may cause false-positive results (15).
- Prior history of laboratory-confirmed Lyme disease
 This is important, as there is no pattern of serological response that can differentiate re-infection from an initial infection with B. burgdorferi (16).

Testing for Lyme disease

Although there are several testing strategies that can assist in making a diagnosis of Lyme disease (**Table 1**), serology is currently the only standardized laboratory testing available. The following describes the different test platforms used for the laboratory diagnosis of Lyme disease. The advantages and limitations of each are presented in **Table 2**.

| Stage of infection | Recommended testing | Specimen type |
|---|---|------------------------------------|
| | strategy^ | |
| Erythema migrans, acute phase (seasonal occurrence and exposure in an endemic area**) | Clinical diagnosis and empirical treatment | None |
| Erythema migrans, acute phase (out of season or no known exposure in an endemic area) | 2-tiered serology [†] – repeat EIA in four weeks if negative; treatment at physician's | Serum |
| endemic area) | discretion NAAI, isolation | Biopsy, plasma |
| Characteristic neurological, cardiac or joint | 2-tiered serology [†] | Serum |
| involvement | NAAT | Synovial or cerebrospinal fluid |
| Persistent symptoms following recommended treatment | None | None |

Table 1: Laboratory testing approaches for Lyme disease (9)

* Tests, such as nucleic acid amplification test (NAAT) or bacterial isolation, are not frequently requested or performed.

** Endemic areas are localities where blacklegged ticks are established and *B. burgdorferi* cycles of transmission are

maintained. Enzyme immunoassay (EIA) followed, where appropriate, by confirmatory Western blots using diagnostic kits licensed in Canada.

Serology testing

The Canadian Public Health Laboratory Network recommends a two-tiered approach to Lyme disease testing, consisting of a sensitive enzyme immunoassay (EIA) followed, if positive or equivocal, by a specific Western blot test (9). The rationale for this approach is that the overall sensitivity and specificity are maximized when these tests are performed in sequence.

The immune response to *B. burgdorferi* infection begins with the appearance of IgM antibodies, usually within two weeks of a tick bite (17). These antibodies may persist for months or even years despite effective antimicrobial therapy (18). Following that IgM response, IgG antibodies develop in most patients, typically after one month of infection (9).

Serology provides a snapshot of the immune status of the patient at the time of the specimen collection. For instance, if Lyme disease is suspected on the basis of symptoms but early serological testing is negative; followup testing on a convalescent sample is recommended (9). The two most commonly performed serological tests are detailed below.

Enzyme immunoassay (EIA)

An enzyme immunoassay is used as a screening test to detect IgM and/or IgG antibodies in serum that are directed against the bacterium that causes LYME DISEASE. Commercial kits, such as an enzyme-linked immunosorbent assay, rely on the use of whole-cell preparations of *B. burgdorferi* (1) and/or recombinant antigens (19) (e.g. C6 peptide). The format of the assay allows the simultaneous screening of a relatively large number of samples. While most enzyme immunoassays are highly sensitive, they may lack specificity (i.e. false positives can occur as a result of other medical conditions).

Western blot (WB)

The Western blot test is used as the corroborative test, and it has greater specificity than the enzyme immunoassay (11, 20). It detects antibodies in serum that are directed against electrophoretically separated antigen extracts and recombinant antigens native to *B. burgdorferi* (21). Commercial kits are used to test for antibodies to individual genospecies of *Borrelia* (12) and to differentiate IgM from IgG antibodies. A positive Western blot result is required to confirm exposure to *B. burgdorferi* (22), and seroconversion from IgM to IgG Western blot antibodies provides definitive evidence of a recent infection (9).

| Test | Advantages | Limitations | |
|---|---|---|--|
| Enzyme immunoassay High sample throughput and relatively easy to perform. Generates objective numerical values compared with other subjective measures (e.g. immunofluorescent assays). | High sample throughput and relatively | Lower sensitivity in early stage Lyme disease. | |
| | easy to perform.Generates objective numerical values | There is variation in the sensitivities and specificities of the different commercial kits available in Canada. | |
| | • Antibodies in the serum of patients with autoimmune disorders, Epstein-Barr virus infection, bacterial endocarditis, syphilis, other spirochetal infections, anaplasmosis or <i>Helicobacter pylori</i> infection may cause false-positive results. | | |
| | Some tests cannot discriminate between antibodies produced against North American versus European/Asian genospecies of <i>Borrelia</i>. | | |
| | • Genotype of <i>B. burgdorferi</i> may reduce sensitivity in early Lyme disease. | | |
| | | Cannot differentiate a previous infection from a re- infection with <i>B. burgdorferi</i>. | |

| Table 2: Laboratory | y tests for L | .yme disease | and their adv | vantages and | limitations |
|---------------------|---------------|--------------|---------------|--------------|-------------|
| | | | | | |

| Test | Advantages | Limitations | |
|--|--|---|--|
| Western blot High specificity such that these tests be used to rule out other etiologic ag Able to determine reactive immunogl classes (IgG vs. IgM) and help different to the second sec | High specificity such that these tests can be used to rule out other etiologic agents. Able to determine reactive immunoglobulin classes (IgG vs. IgM) and help differentiate | Interpretation of results is subjective (e.g. scoring band position and intensity) for Western blot assays that do not use an automated reader. Significant cross-reactivity occurs among European | |
| | early from longer-standing infections. | genospecies. | |
| | | LgM antibodies are inherently cross-reactive, which may lead to false-positive results. | |
| | | False-negative IgG Western blot results may occur early in the course of infection or as a result of antibiotic treatment. | |
| SUPPLEMENTAL LABORATORY TESTS (Not routinely performed) | | | |
| Bacterial isolation | Highly specific and could be useful for determining infecting genotypes of <i>B. burgdorferi</i> . | Collection of specimens can be invasive. | |
| | | Relatively low sensitivity, expensive, labour-intensive and long incubation period required for results. | |
| Nucleic acid amplification testing (NAT) | Able to detect <i>B. burgdorferi</i> DNA after antibiotic treatment has started, therefore | Poor sensitivity due to low bacterial load in some clinical samples. | |
| | able to distinguish an ongoing infection from persistent symptoms due to an immunologic mechanism. | Lack of standardization with respect to target genes. | |
| | Borrelia DNA can be detected in EM lesions before the appearance of serum antibodies and without the delay associated with bacterial isolation. | | |

Two-tiered algorithm for the laboratory diagnosis of Lyme disease

The two-tiered approach to testing is illustrated in **Figure 1**. The first tier involves the use of an EIA. If this EIA test is negative, WB testing is not indicated. If symptoms persist, the EIA test can be repeated on a convalescent sample collected 3–6 weeks later. If the EIA is positive or equivocal, the second tier or corroborative Western blot assay is performed. In early infections (i.e. symptoms for less than six weeks), both the IgM and IgG Western blot tests are performed; however, if the patient has had symptoms for more than six weeks, only the IgG Western blot assay is performed.





The final result of serological testing is considered positive only when the EIA is reactive (positive or equivocal) and the WB is also positive (**Table 3**). This two-tiered system maximizes the sensitivity and specificity of the assays and increases the likelihood of observing a seroconversion (from IgM to IgG) that is evident in most *bona fide B. burgdorferi* infections (1, 17, 21, 23).

Table 3: Interpretation of Western blot results (in conjunction with an equivocal or positive EIA)

| Western blot result | Interpretation |
|---|---|
| Both IgM and IgG Western blots negative | Result not consistent with a <i>B. burgdorferi</i> infection; however, if symptoms persist submit a follow-up sample 3–6 weeks later. |
| Only IgM Western blot positive* | Potentially a false-positive result if this is NOT an acute case (i.e. < 6 weeks post onset of symptoms). |
| Only IgG Western blot positive** | Result consistent with an infection with <i>B. burgdorferi</i> of greater than 4 weeks' duration. |
| Both IgM and IgG Western blots positive | Result indicates recent or previous infection with <i>B.</i> burgdorferi. |

IgM positive WB – 2 of 3 significant bands present.

^{*} IgG positive WB – 5 of 10 significant bands present (22).

Supplemental laboratory tests to detect B. burgdorferi

Bacterial isolation

The recovery of viable *B. burgdorferi* from clinical specimens is accomplished by incubating the sample in specialized culture medium. Although this test remains the "gold standard" for diagnosis of Lyme disease(7), the procedure is expensive to perform, lacks clinical sensitivity and is prone to contamination (24). The greatest practical limitation is that cultures can require up to eight weeks of incubation because of the small number of viable organisms present in many specimen types (17). These factors reduce the clinical applications of this test and restrict its use to research studies (9).

Nucleic acid amplification testing (NAAT)

Nucleic acid amplification testing (NAAT) has been used to decrease turnaround times for Lyme disease diagnostic results (25, 26). Several formats of polymerase chain reaction (PCR) testing (i.e. nested, real-time or quantitative) are used to amplify a variety of *Borrelia*-specific genetic targets in clinical specimens (27). Positive results are most frequently seen in the early phase of the disease (28). The sensitivity of polymerase chain reaction on cerebrospinal fluid (CSF) is low or variable and therefore of limited usefulness in evaluating patients with neurological signs (1, 21). Although these assays can identify an infection sooner than serological testing (25, 27), their use is restricted to research studies at the present time (8, 9).

Challenges associated with diagnostic testing for Lyme disease

Physicians and laboratory scientists have concerns regarding results reported by some private laboratories that are inconsistent with results obtained by Canadian public health laboratories. A number of private laboratories may not be using sufficiently validated tests or interpretation criteria. The use of assays that do not have adequately established accuracy and have not been sufficiently validated may result in the reporting of false-positive results.

Some of these tests include capture assays for antigens in urine, immunofluorescence staining, cell sorting of cell wall-deficient or cystic forms of *B. burgdorferi*, lymphocyte transformation tests (29) and a new culture method for serum (30).

Currently, not following the two-tiered algorithm (e.g. by performing a Western blot alone or after the EIA is negative) can increase the frequency of false-positive results. This in turn could lead to possible misdiagnosis and unnecessary treatment (1). Clinicians should have an understanding of the current common misconceptions

about LYME DISEASE(31) and know the best laboratory practices to diagnose it (1); this would facilitate informed discussions with patients who have questions and concerns.

Future developments

As mentioned, there are a wide variety of diagnostic tests available to assist in the diagnosis of Lyme disease (21, 32), and considerable debate exists concerning the accuracy and reliability of some of these tests (32–34). At this time there is no single laboratory test that is 100% sensitive and specific for the confirmation of Lyme disease. This is further complicated by the fact that not all individuals who are infected with *B. burgdorferi* present in the same way. Improvements in diagnostic test platforms are a priority. The search for "biomarkers" of Lyme disease that do not rely on serological response is ongoing (35). Variations of the two-tiered approach, which typically reduce or eliminate the use of WBs, are being evaluated (36–39) and may help simplify test interpretation and improve sensitivity in early Lyme disease.

One of the biggest challenges to evaluating new diagnostic approaches (40) in Canada is the lack of serum panels or a collection of well-characterized samples from individuals with confirmed *B. burgdorferi* infection. It is also imperative that we gain a full understanding of the genotypes of *B. burgdorferi* that are infecting Canadians (41) and establish whether the diagnostic tests currently in use can detect all of them with comparable sensitivity.

The Agency's National Microbiology Laboratory plans to work with diagnostic laboratories across Canada to review current Lyme disease diagnostic practices and quality assurance systems, and to evaluate the need for enhanced internal and external proficiency testing. In the longer term, the Agency also plans to 1) review and update the existing laboratory guidelines for Lyme disease, 2) determine and compare test performance characteristics of all EIA and Western blot platforms currently in use in Canada, 3) evaluate "new" diagnostic platforms on a high priority and ongoing basis and 4) initiate the process of development of a robust serum panel for use in evaluation of new assays (42). This work will advance quality diagnostic testing for Lyme disease in Canada.

Conclusion

The incidence of Lyme disease is increasing in Canada. In persons with erythema migrans rash and a reliable history of exposure to blacklegged ticks, testing is not required and treatment can be started empirically. The clinical assessment of patients in the disseminated phases of Lyme disease can be further supported by laboratory testing.

At this time, there is no perfect laboratory test for Lyme disease; however, the two-tiered serological approach provides the most sensitive and specific results to date. Improper use of serologic tests or the use of diagnostic tests or interpretative criteria that have not been fully validated may lead to misdiagnosis and unnecessary antibiotic treatment. Future efforts will focus on standardization of testing, and the development and evaluation of new diagnostic approaches to optimize the detection of Lyme disease.

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Conflict of interest

The authors have no conflicts of interest to declare.

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